

### **Original Article**

### CHANGES IN HAEMATOLOGICAL PARAMETERS OF CLARIAS GARIEPINUS EXPOSED TO CENTURY PLANT (*Agave americana*) LEAF DUST

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#### ABSTRACT

The effect of sub-lethal concentrations (0.1250, 0.0625, 0.0313, 0.0156, 0.0078 mg/L and Control) of *A. americana* leaf dust on haematological parameters of *C. gariepinus* was investigated, using static renewable bioassay system for 28-day period. There was significant difference (p<0.05) in the monitored physico-chemical parameters (hydrogen ion concentration (pH), total alkalinity, dissolved oxygen (DO) and free carbon (iv) oxide (CO<sub>2</sub>)), however, temperature was not significantly different (p>0.05) in aquaria with *A. americana* compared to the Control. The packed cell volume, red blood cell counts, haemoglobin, and mean corpuscular haemoglobin concentration were significantly depleted (p<0.05) whilst, white blood cell counts significant increased (p<0.05) as the concentration of *A. americana* leaf dust increased. The implication of these findings revealed that the leaf dust of *A. americana* have negative effects on the test fish which may help explain the decline in wild fishery resources, especially, in areas where the plant is used for obnoxious fishing practices.

**Keywords:** African catfish,Blood Parameters, *Clarias gariepinus*, Medicinal plant, Sublethal effect,

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## INTRODUCTION

The aquatic ecosystem like the terrestrial environment, is continuously subjected to changes in quality that are due to the introduction of substances of diverse characteristics arising from man's cultural activities (Oluah, 2001). The accumulation of toxicants in an aquatic environment can result in reduced reproductive capabilities, alter growth rates and reduce ability to withstand variations in pH, temperature and dissolved oxygen (Adamu et al., 2008).

Majority of herbal plants and their products have been used as natural alternatives for treatment and management of various diseases including hepatic disorders (Stickel et al., 2005), as pesticides (Maikai et al., 2008), as molluscides (Azare et al., 2007) and piscicides (Tiwari and Singh, 2004). According to FAO (1991), more than 60,000 plant species are used for various purposes all over the world. Istvan (2000) reported that plants are virtually an inexhaustible source of biological active substances. The usefulness of plants for piscicidal and medicinal purposes has been reported (Adewole et al., 2002; Adamu, 2009; Akobundu, 1987) and the use of toxic plants to harvest fish is common practices worldwide (Ugwemorubong et al., 2009). Kawazu,(1972) reported that piscicidal plants have been used by traditional societies all over the world as a means of catching fish in small bodies of water. Piscicidal plants

like *Blighia sapida, Kigelia africana, Tetrapleura tetraptera, Raphnia vinifera, and Parkia biglobosa* are commonly used by fisher folks to harvest fishes (Fafioye *et al.*, 2004).

The eradication of wild fishes in the culture ponds before stocking of desired species is an important step in pond management as the former compete and/or prey upon the latter. Consequently, the control and eradication of unwanted fishes in the pond requires effective piscicides which are usually not easily accessible; farmers use synthetic compounds including malachite green (Ayotunde et al., 2011). Different species of plants employed as piscicides have different effects, depending on the species of fish targeted. According to Yadav (2000), A. americana has been used by fishermen in catching fish. It contains different active ingredients such as nicotine, pyrethrum, rotenone, resin, tannins and saponins (Wang and Huffman, 1991).

Agave americanacommonly known as century plant, maguey or American aloe belongs to the Family Asparagaceae. The plant is originally native to Mexico, Arizona and Texas. They are cultivated worldwide as an ornamental plant. The plant has become naturalized in many regions including the West Indies, parts of Africa and South America.

Fish have been the most popular choice as test organisms because they are presumably the best understood organisms in the aquatic environment

(Buikema et al., 1982) and also due to their importance to man as a protein source (Kime et al., 1996). The African catfish (*Clarias* gariepinus) are freshwater fishes found in the tropical regions of West African; therefore this fish may be one of the non targeted organisms that may be affected by the use of A. americana. In order to ascertain the effect of the plant piscicide on the health status of the test fish before death, haematological been recommended studies have (Kori-Siakpere et al., 2005). Such studies have generally been used as an effective and sensitive index to monitor physiological and pathological changes in fishes (Iwama et al., 1976). According to Babatunde et al., (1992), changes in the constituents' component of blood sample when compared to the blood profile could be used to interpret the metabolic state of fish and its state of health. This present study investigated the sub-lethal effect of A. americana on the haematological indices (packed cell volume, red blood haemoglobin, cell counts, mean corpuscular haemoglobin, and mean corpuscular haemoglobin concentration and white blood cell counts) of the African catfish (*Clarias gariepinus*) after 28-day exposure period.

# MATERIALS AND METHODS

The juveniles of *Clarias gariepinus* of mean weight and length of  $30\pm0.20$ mgand  $17\pm0.40$ cm respectively of mixed sex were purchased from Renajj Fish Farms, Rayfield, Jos, Plateau State, Nigeria. The juveniles were acclimatized for 14 days in 60L capacity circular tanks in the Departmental Laboratory, Department of Zoology, University of Jos. During the acclimatization and trial periods, the fish were fed with coppens feed. Similarly, during the acclimatization and trial periods, 50% of water was siphoned daily in order to remove leftover feed and faecal matter. The water was thereof replaced with fresh 50% dechlorinated municipal tap water that was stored in laboratory condition.

The leaves of A. americana were collected from the University main Campus. Identification of the plant was aided by the technical staff of the Department of Botany. The leaves were air dried before being ground into powder and sieved with 30µm mesh The concentrations size. of А. americana leaf dust used was achieved after series of preliminary and acute toxicity tests. 0.125, 0.0625, 0.0313, 0.0156, 0.0080 and 0.000mg/L were concentrations used for the exposure. During the 28-day experimental some physico-chemical period, parameters such as pH, dissolved oxygen (DO), free carbon (iv) oxide  $(CO_2)$ , total alkalinity and temperature (°C) were monitored using standard procedures of APHA, (1998) weekly with the exception of temperature that was determined daily. At the end of the 28-day experiment, blood samples collected from all the were experimental fishes ethylene in diamine tetra acetic acid (EDTA) test for the determination tubes of haematological indices as described by Blaxhall and Daisley (1973).

The data from the physico-chemical and haematological parameters were subjected to analysis by calculating means, standard error and Analysis of Variance (ANOVA) and student's t-test at 5% probability level.

#### RESULTS

The mean values of the monitored physico-chemical parameters of the test and control media are presented in Table 1. The pH and dissolved oxygen decreased significantly (p<0.05) while free carbon (iv) oxides and total

significantly alkalinity increased (p < 0.05) as the concentrations of the plant leaf dust increased. The haematological indices of *Clarias* gariepinus exposed to sub-lethal concentrations of A. americana leaf dust and the Control after the 28 days exposure period are presented in Figures 1 to 6. The mean value of packed volume cell (PCV). haemoglobin (Hb), mean corpuscular haemoglobin (MCH), mean corpuscular haemoglobin concentration (MCHC) and red blood cells (RBC) were found to significantly (p < 0.05) decrease as the concentration of *A. americana* leaf dust increased while that of white blood cells (WBC) showed significant (*p*<0.05) increase as the concentration of A. americana leaf dust increased.

Table1: Mean (standard error) physico-chemical parameters for sub-lethal bioassay of *A. americana* leaf dust on *Clarias gariepinus* for 28 days.

Parameters	A. americana leaf dust concentration (mg/L)					
	0.0000	0.0080	0.0156	0.0313	0.0625	0.1250
Temperature(°C)	22.50	22.50	22.50	22.50	22.50	22.50
	(0.10)	(0.10)	(0.10)	(0.10)	(0.10)	(0.10)
рН	7.09	6.77	6.70	6.90	6.66	6.22
	(0.03)	(0.41)	(0.15)	(0.18)	(0.55)	(0.57)*
Dissolved oxygen(mg/l)	5.85	5.30	4.85	4.62	4.45	4.30
	(0.05)	(0.10)	(0.23)	(0.38)	(0.29)*	(0.09)*
Free carbon (iv) oxides (mg/L)	3.57	3.75	4.00	4.15	4.32	4.60
	(2.38)	(0.38)	(3.30)	(2.10)	(0.05)*	(0.04)*
Total alkalinity (mg/L)	6.90	8.62	10.63	11.92	14.40	15.97
	(0.05)	(1.58)	(3.27)*	(4.40)*	(6.16)*	(7.02)*

\* significant difference *p*<0.05 compared to the control.

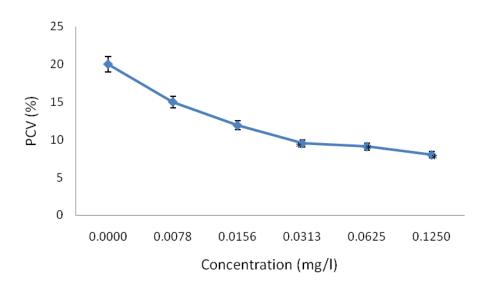


Fig 1: Packed cell volume of *C. gariepinus* exposed to concentrations of *A. americana* leaf dust for 28 days. Vertical bars represents standard error, \* = p < 0.05 compared to control

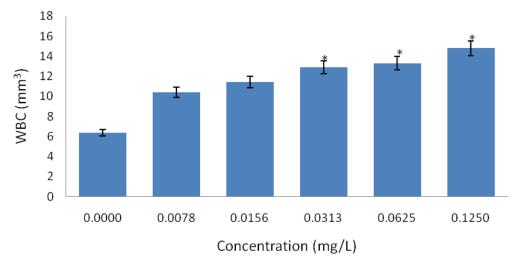


Fig 2: White Blood Cell level of *C. gariepinus* exposed to concentrations of *A. americana* leaf dust for 28 days. Vertical bars represents standard error, \* = p < 0.05 compared to control

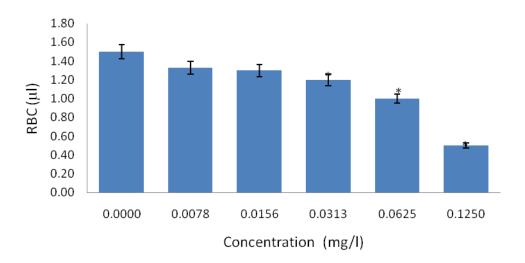


Fig 3: Red Blood Cell level of *C. gariepinus* exposed to concentrations of *A. americana* leaf dust for 28 days. Vertical bars represents standard error, \* = p < 0.05 compared to control

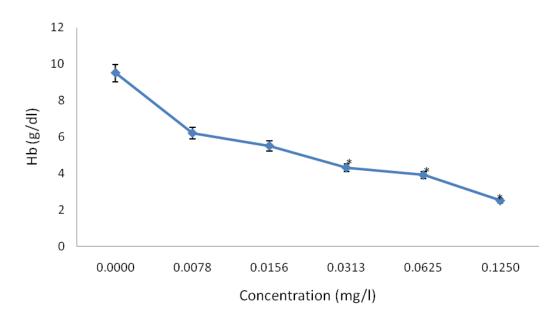


Figure 4: Haemoglobin level of *C. gariepinus* exposed to concentrations of *A. americana* leaf dust for 28 days. Vertical bars represents standard error, \* = p < 0.05 compared to control

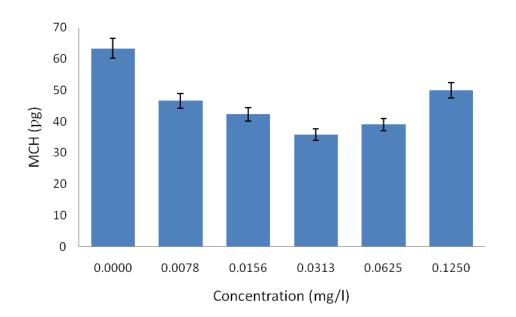


Fig 5: Mean corpuscular haemoglobin level of *C. gariepinus* exposed to concentrations of *A. americana* leaf dust for 28 days. Vertical bar represents standard error

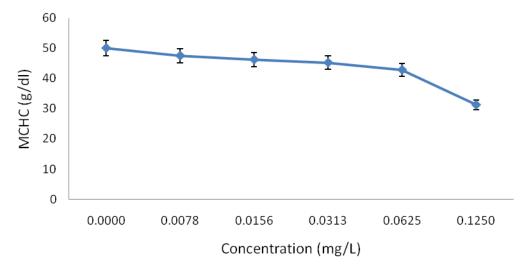


Fig 6: Mean corpuscular haemoglobin concentration level of *C. gariepinus* exposed to concentrations of *A. americana* leaf dust for 28 days. Vertical bar represents standard error

#### DISCUSSION

Physico-chemical parameters such as temperature, dissolved oxygen, free carbon (iv) oxide, pH, alkalinity are paramount to the many factors which affect fish health, growth and reproduction (Camus *et al.*, 1998). In this study, the monitored parameters were noted to be significantly different from the Control except for temperature. The decline in pH with time may be due to the production of acidic metabolites (Delyan *et al.,* 1990) by the plant material in water. Noga (1996) recommended pH range of 6.5 to 8.5 for fresh water fishes, the value ofpH in the two highest concentrations of the plant leaf dust were found to be lower than the recommended value. Prasad *et al.* (1995) reported that the reduction in dissolved oxygen content in a bioassay media as toxicant concentration increased may be due to antioxidant property of the toxicant. Similarly, free carbon (iv) oxide content increased with increase in the concentration of the toxicant.

The sub-lethal toxicity test carried out showed that A. americana leaf dust caused significant changes in the hematological indices of *C. gariepinus*. The changes in the values of the haematological indices of the test fish is similar to that reported by Ayotunde et al. (2011). The packed cell volume, haemoglobin and erythrocyte counts indicators are good of oxygen transportation capacity of fish thus making it possible to establish relationship with the oxygen concentration available in the habitat and the health status of the fish (Lamas et al., 1994). On the other hand the white blood cells confer protection against infectious agent caused by microbial and chemical factors (Gusmao et al., 2007).

The significant reduction in PCV, Hb, RBC, could be indication of severe anaemia caused by destruction of erythrocytes (Omoniyi *et al.*, 2002) or haemo-dilution (Adeyemo, 2005), resulting from impaired osmoregulation across the gill epithelium as there was significant decrease in dissolved oxygen level. PCV is used to determine the ratio of plasma to corpuscles in the blood as well as the oxygen-carrying capacity of the blood (Larsson et al., 1985). Adamu and Audu (2008) reported that the significant decrease in PCV may be attributed to gill damage and/or osmoregulation causing impaired haemodilution. anaemia and Haemoglobin is the oxygen-carrying component in the blood of fish and its concentration can be used as good indicator of anaemia (Blaxhall and Daisley, 1973). The decrease in Hb corresponds with the decrease in dissolved oxygen; an indication that the decrease in haemoglobin resulted in haemodilution. The Hb values fall lower than the range reported for catfish (Iheukwumere et al., 2002; Ayotunde et al., 2011). The reduction may be due to increased rate of breakdown of red blood cells and/or reduction in the rate of formation of red blood cells (Mossa, 2004) which may probably have been caused by the dust. Haemoglobin plant leaf concentration and packed cell volume values are directly correlated to erythrocytes count which may be due to the synergistic linkage of the blood cells. The increase in the white blood cells may have been induced as protection against disease and improving the health mechanism of the fish in the stressed condition.

The MCH and MCHC of the test fish were found to decrease with increasing toxicant concentration. Similar reduction has been observed by

Okomoda et al. (2010) in C. gariepinus formalin. exposed to The mean corpuscular haemoglobin concentration which is the ratio of the mean haemoglobin concentration is not influenced by blood volume neither by the number of cells in the blood, but can be interpreted incorrectly only when new cells, with a different haemoglobin concentration are released (Tawari-Fufeyin et al., 2008). Gafaar et al.(2010) reported that prolonged reduction in haemoglobin content is deleterious to oxygen transport and degeneration of the erythrocytes could be due to pathological condition in fish exposed to toxicants.

# CONCLUSION

The study revealed that sub-lethal concentrations of *A. americana* leaf dust has deleterious and debilitating effects on the haematological indices of *C. gariepinus* juveniles as it also interferes with the water quality. Therefore, the use of the plant leaf dust should be discouraged.

# REFERENCES

Adamu, K.M. (2009). Sub-lethal effect of tobacco (Nicotiana tobaccum) leaf dust on Enzymatic activities of heteroclarias hybrid of (a Heterobranchus bidorsalis and Clarias gariepinus), Jordan Journal of *Biological Sciences*, 2(4): 151-158.

Adamu, K.M. and Audu, B.S.(2008). Haematological assessment of the Nile tilapia *Oreochromis niloticus* exposed to sublethal concentrations of Portland cement powder in solution, *International Journal of Zoological Research*, 4(1): 48-52.

Adamu, K.M., Audu, B.S. and Audu,<br/>E.L.(2008).ToxicityandHistopathological effects of Portland<br/>cement powder in solution on the<br/>structure of the gill and liver tissues of<br/>the Nile tilapia, *Oreochromis niloticus:*<br/>A microscopic study, *Tropical*<br/>*Freshwater Biology*,17(1): 25-36.

Adewole, A.M., Faturoti, E.O., Oladeinde, O.F. and Ayelaagbe, O.O. (2002).A survey of some indigenous fish phytotoxic plants in Ibadan, South Western Nigeria. Book of Abstract of the 1<sup>st</sup> Annual Conference of the Zoology Society of Nigeria.123-140.

Adeyemo, O.K. (2005). Haematological and histopathological effects of cassava mill effluent in *Clarias gariepinus*. *African Journal of Biomedical Research*, 8: 179-183.

Akobundu, I.O. (1987). Weed science in the tropics. *Principles and Practices*. John Wiley and sons, Chichester, New York.33-38.

APHA, (1998).Standard methods for the examination of water and waste water, 20<sup>th</sup> edition (Revised edition), American Public Health Association NY USA, 1076pp.

Ayotunde, E.O., Offem, B.O. and Bekeh, A.F.(2011). Toxicity of *Carica papaya* 

lim: Haematological and piscicidal effect on Adult catfish (*Clarias gariepinus*). *Journal of Fisheries and Aquatic Science*, 6(3): 291 – 308.

Azare, B.A., Okwute, S.K. and Kela, S.L. (2007). Molluscicidal activity of crude water leaf extracts of *Alternanthera sesselis* on *Bulinus* (phy) *globosus. African Journal of Biotechnology*,6: 441-444.

Babatunde, G.M.,Fajimi, A.O. and Oyejide, A.O. (1992).Rubber seed oil versus Palm oil in broiler chicken diet.Effect on performance, nutrient digestibility, haematology and carcass characteristics. *Animal Feed Science Technology*,35: 133-146.

Blaxhall, P.C. and Daisley, K.W. (1973). Routine haematological methods for use with fish blood. *Journal of Fish Biology*, 5: 771-781.

Buikema, A.L (Jr).Niederlehner , B. R and Cairns, J.(Jr). (1982). Biological monitoring: Part IV- *Toxicity Testing Water Resource*, 16: 239-262.

Camus, A.C., Burrow, R.M., Hemstreet, W.G., Thure, R.L. and Hawke, J.P. (1998). Aeromonas Bacterial infections- Motile*Aeromonas septicemia. Southern Regional Aquaculture Centre Publication*, No.478.

Delyan, U., Harder, H. and Hopner, T.H. (1990).Hydrocarbon biodegradation in sediments and soils. A systematic examination of physical and chemical conditions part 11.pH values. *Wisseschaft Technik Scientific Technology*, 43: 337-342.

Fafioye, O.O., Adebisi, A.A. and Fagade, (2004). Toxicity S.O. of Parkia biglobosa Raphnia vinifera and extracts Clarias on gariepinus African Journal juveniles. of *Biotechnology*, 3(11):627-630.

FAO.(1991). *Energy for sustainable rural development projects* – Training materials for Agricultural Planning.....?

Gafaar, A.Y., El-manakhly, E.M.. Soliman, M.K., Soufy, H., Monas, Z., Mohammed, S.G. and Hassan, S.M. (2010). Some pathological, biochemical and Haematological investigation on Nile tilapia (*Oreochromis niloticus*) chronic exposure following to edifenphos Journal of pesticide. American Science, 6(10) 542-551.

Gusmao, A. E., Da Costa, S.E., Tavares-Dias, M.G., Cruz de Menezes, G.C., Suely-Melo, C.E., Da Silva, E.S.N., Rebelo, D.I., Roubach, R.E., Akifumi, E.O., Daniel, J.I.F. and Luiz, J.M.(2007). Effect of high levels of dietary vitamin C on the blood response of matrinxa, *Brycona mazonicus. Comparative Biochemistry and Physiology*, 147B: 383-388.

Iheukwumere, F.C, Okoli, I.C. and Okeudo, N.I.(2002). Preliminary studies on raw napolena imperials seed as food ingredients: Haematology and serum biochemistry, carcass and oxygen weight of weaner rabbits. Audu *et al*.

*Tropical Animal Production Invest.,*5: 219 – 227.

Istvan,U.(2000).Semi-naturalProducts and related substances asalleged botanical pesticides.Management Sciences, 56 (8) 703-705.

Iwama, G.K., Greer, G.L. and Larkin, D.A. (1976). Changes in some haematological characteristics of Coho Salmon (*Oncorhychus kisutch*) in response to acute exposure to dehydroabietic (DHAA) acid at different exercise levels. Journal of Fisheries Research, 33: 285-289.

Kawazu, K. (1972). Active constituents of piscicidal plants. *Yuukigoseikagaku,* 30: 615-628.

Kime, D.E., Ebrahimi, M., Nysten, K., Roelantis, I., Rurangwa, E., Moore, H.D.M. and Ollevier, F. (1996). Use of computer-assisted sperm analysis (CASA) for monitoring the effects of pollution on sperm quality of fish, application to the effects of heavy metals. *Aquatic Toxicology*, 36:223-237.

Kori-siakpere, O., Ake, J.E.G. and Idoge, E. (2005). Haematological characteristics of the African snakehead, *Parachanna obscura*. *African Journal of Biotechnology*,4 (6):527-530.

Lamas, J., Santos, Y., Bruno, D.W., Toranzo, A.E. and Anadon, R.(1994).Non specific cellular responses of rainbow trout to *vibrio*  *anguillarium* and its extracellular products (ECPs). *J. Fish Biol.*,45 (5): 839-854.

Larsson, A., Haux, C. and Sjobeck, M. (1985).Fish physiology and metal pollution result and experience from laboratory and field studies. *Ecotoxicol. Environ. Saf.,*9: 250-281.

Maikai, V.A., Kobo, P.I. and Adaudi, A.O. (2008). Acute toxicity studies of aqueous stem bark extract of *Ximenia americana. African Journal of Biotechnology*, 7: 1600-1603.

Mossa, A.H. (2004). Genotoxicity of pesticides. PhD Thesis. Pesticides chemistry and toxicity, Faculty of Agriculture, Damantiour Alexandria University.....?

(1996). *Fish* Noga, E.J. Disease: Diagnosis and Treatment Mosby Yearbook, Incoporated. Weslin industrial Drive: St. Louis, Missouri,367pp.

Okomoda, J., Ayuba, V.O. and Omeji, S. (2010). Haematological changes in *Clarias gariepinus* (Burchell, 1822) fingerlings exposed to acute concentrationof formalin, *Production Agriculture and Technology*,6(1): 92-101.

Oluah, N. S. (2001). The effect of sublethal cadmium on the haematology of the freshwater catfish, *Clarias gariepinus* (Pisces: Clariidae), *Journal of Science, Agriculture, Food Technology and Environment*, 1: 9 - 14. Omoniyi, I., Agbon, A.O. and Sodunke, S.A. (2002). Effect of lethal and sublethal concentration of Tobacco (*Nicotiana tobaccum*) leaf dust extract on weight and haematological changes in *Clarias gariepinus* (Burchell). *Journal of Applied Sciences and Environmental Management*, 6(2): 37-41

Prasad, K., Laxdal, V.A., Ming, Y.U. and Raney, B.L. (1995). Antioxidant activity of Allicin and active ingredient of Garlic. *Molecular and Cellular Biochemistry*, 1489 (2) 183-189.

Tawari-Fufeyin, P, Igetei, J. and Okoidigun, M.E. (2008). Changes in the catfish (*Clarias gariepinus*) exposed to acute cadmium and lead poisoning. *Bioscience Research Communication*, 20(5): 271-276.

Tiwari, S. and Singh, A.J. (2004). Changes in some biochemical parameters in the liver and muscle of *Colisa fasciatus* due to toxicity of ethanolic extract of *Nerium indicum* latex. *Natural Product Radiance*,8: 48-54.

Ugwemorubong, U.G. Obomanu, F. and Oveh, O.D. (2009). Enzymes in selected tissues of catfish Hybrid exposed to Aqueous extracts from *Lepidagathis alopecuroides* leaves. *Electronic Journal of Environmental Agriculture and Food Chemistry*,1579- 4377.

Wang, S. and Huffman, J.B. (1991).Botanochemicals: Supplements to petrochemicals. *Economy Botany*, 35(4)369-382.

Yadav, S. (2000). Survey of capture fisheries in the Koshi River Basin. *Fisheries Training Centre*, Janakpur. Dhanusha, Nepal. *Food and Agricultural Organization (FAO) Corporate Document Repository.*